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Isolated cortical computations during delta waves support memory consolidation

Ralitsa Todorova and Michaël Zugaro*

Delta waves have been described as periods of generalized silence across the cortex, and their alternation with periods of endogenous activity results in the slow oscillation of slow-wave sleep. Despite evidence that delta waves are instrumental for memory consolidation, their specific role in reshaping cortical functional circuits remains puzzling. In a rat model, we found that delta waves are not periods of complete silence and that the residual activity is not mere neuronal noise. Instead, cortical cells involved in learning a spatial memory task subsequently formed cell assemblies during delta waves in response to transient reactivation of hippocampal ensembles during ripples. This process occurred selectively during endogenous or induced memory consolidation. Thus, delta waves represent isolated cortical computations tightly related to ongoing information processing underlying memory consolidation.

ost of our time spent asleep is dominated by slow oscillations (0.1 to 1 Hz), when cortical neurons synchronously alternate between a depolarized (up) state associated with high levels of endogenous activity and a hyperpolarized (down) state when neurons remain silent (1). Delta waves are large deflections of the local field potential (LFP) that correspond to the down states of the slow oscillation and are thus considered periods of generalized cortical silence. The slow oscillation plays a causal role in memory consolidation (2-5), in particular by orchestrating an information flow between the hippocampus and the neocortex (6). Indeed, delta waves tend to occur in close temporal proximity to hippocampal ripples (7), which are instrumental for memory consolidation (8, 9). Hippocampal replay of awake activity (10), biased by inputs from sensory cortices (*11*, *12*), initiates reactivation of prefrontal cortical cell assemblies (*13*, *14*) just before the occurrence of a delta wave (*7*). Cortical synaptic plasticity subsequently takes place during network reorganization early in the following up state (*15*, *16*) and during the massive calcium entry accompanying the ensuing sleep spindle (*17–19*). This hippocampo-cortical dialogue (*20–22*) is instrumental for memory consolidation (*5*). However, the incursion of generalized silence (delta wave) precisely between periods of information exchange and periods of network plasticity remains puzzling.

We recorded prefrontal cortical activity in nine rats during slow-wave sleep (5). Consistent with previous reports, most delta waves were accompanied by neuronal silence. Yet, occasionally, spikes did occur during delta waves (Fig. 1A), and when considering cumulative spiking activity over all recorded delta waves, unexpected residual activity appeared at the peak of the waves (Fig. 1, B and C) (spike waveforms recorded during delta waves were not distinguishable from spike waveforms

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Fig. 1. Delta spikes.

(A) Example of a nonsilent delta wave. Colored curves indicate LFPs recorded from the medial prefrontal cortex (color: recording channel). Colored vertical ticks indicate spikes emitted by simultaneously recorded prefrontal units (color: channel from which the unit was recorded). Dashed lines indicate the beginning and end of delta waves. A delta spike (black circle) occurs during the second delta wave, when the rest of the network remains silent. Black calibration bars: 0.5 s (horizontal); 1 mV (vertical). mPFC, medial prefrontal cortex. (B) Mean perievent time histogram of the normalized firing rate of



prefrontal units centered on delta waves (top curve: mean field event). The dashed white line indicates residual activity during delta waves. (**C**) Time distribution of the spikes emitted by each prefrontal neuron closest to each delta wave. The large peak at ~100 ms corresponds with activity in the up state. The smaller peak consists of spikes occurring during delta waves. The dashed line indicates the 15-ms upper threshold used to define delta spikes in subsequent analyses (all results were confirmed by using ±30-ms time windows).

(**D**) (Left) Number of units that discharged in a given proportion of delta waves (gray curve: log-normal fit with the same mean and variance as the data; error bars: 95% confidence intervals from bootstrapped data). No unit fired in 0% of the delta waves. (Right) Number of delta waves containing a given number of delta spikes. (**E**) No difference in average waveforms between silent (black: n = 101,161) and nonsilent (blue: n = 12,205) delta waves (Monte Carlo test, P > 0.05).

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recorded outside delta waves; fig. S1). On closer examination, neuronal activity occurred consistently in a substantial fraction of delta waves (12%), wherein one or a few neurons remained active while the rest of the population became silent (Fig. 1D). We call this unexpected persisting activity "delta spikes." To investigate whether delta spikes were restricted to a particular subset of neurons, we counted the number of delta waves in which each unit emitted one or more spikes. As it happened, every single recorded unit fired during delta waves, suggesting instead that persisting firing may actually constitute a widespread phenomenon (Fig. 1D and fig. S2).

We then wondered whether delta spikes tended to occur in specific delta waves with distinctive characteristics. We thus compared delta waves in which we did or did not detect cortical spikes and found no significant difference between the two groups in terms of waveform (Fig. 1E), duration, timing (fig. S3), depth (fig. S4), decreased gamma power, or coupling with hippocampal ripples and thalamocortical spindles (fig. S5). This suggests that spikes could take place during virtually all delta waves but may remain undetected given the limited number of recorded neurons relative to the entire population (fig. S6). We thus hypothesize that firing during delta waves might be an overlooked phenomenon manifested in possibly all delta waves.

These findings indicate that during any given delta wave, the cortical network becomes

silent except for a small but ever-changing minority of cells. The most parsimonious explanation would be that delta spikes constitute random activity reflecting imperfect coordination in the cortical alternation between the up and down states. Yet, an alternative possibility is that this activity serves a welldefined computational function. A hallmark of cortical computation is the emergence of cell assemblies. We thus tested for the presence of recurring coactive cell ensembles, using two complementary approaches. As a first approach, we performed a standard independent component analysis (23), which identified multiple significant components that were active during delta waves (fig. S7. A and B). However, these components were likely to



Fig. 2. Hippocampal ripple activity predicts delta spikes. (**A**) Delta spikes and preceding hippocampal activity. Blue traces indicate LFPs from the medial prefrontal cortex. Colored ticks indicate simultaneously recorded prefrontal (blue) and hippocampal (red) spikes. Black circles indicate delta spikes emitted within ±15 ms of the delta wave peak (shaded area). In the first two delta waves, delta spikes were recorded from the same unit after similar hippocampal activity patterns. HPC, hippocampus. (**B**) Cross-correlations (curves and shaded areas, mean ± SEM; orange, data; gray, time-shifted control) between hippocampal ripple activity (sliding window) and delta spikes (fixed, 0 s). The horizontal orange line indicates the Monte Carlo test: P < 0.05. corr., correlation. (**C**) Enrichment in positive correlations [comparative distribution between data and control in (B)] when hippocampal activity was correlated with subsequent prefrontal

delta spikes. (**D**) Performance of a GLM trained to predict prefrontal activity during delta waves on the basis of preceding hippocampal ripple activity (200-ms window). Delta spikes as well as delta components were significantly predicted by multiple single-unit hippocampal activity (P = 0.0403 for delta spikes, P = 0.0052 for delta components; Wilcoxon rank sum test) but not by global hippocampal drive ignoring cell identity (summed activity, P = 0.2597 for delta spikes and P = 0.3258 for delta components; Wilcoxon rank sum test). **P < 0.01; ***P < 0.001, (Wilcoxon signed-rank tests). (**E**) Object responsivity index for partner (green) versus other (gray) prefrontal units (curves: cumulative distributions; inset: mean ± SEM). Only partner prefrontal units showed positive object responsivity (partner units, P = 0.0162; other units, P = 0.5967; Wilcoxon signed-rank test; partner versus other units, P = 0.0465; Wilcoxon rank sum test). **P < 0.05.

combine multiple smaller but overlapping cell ensembles, given the limited number of neurons active in any given delta wave. We thus performed a second analysis to examine cooperative activity ("peer prediction") (24) among delta spikes, an idiosyncratic property of cell assemblies. This showed that the delta spikes of one neuron could be predicted from the delta spikes of other neurons (fig. S7C). We then asked whether delta spikes were involved in the hippocampo-cortical dialogue underlying memory consolidation. Because delta waves typically take place precisely between hippocampal replay and cortical reorganization for memory consolidation, this hypothesis would be expected to have two implications: (i) Hippocampal activity during ripples should predict which neurons (or which

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spikes

assemblies) are active during the following delta wave, and (ii) this predictive bias should emerge after behavior, and predictable cortical cells should be involved in the reactivation of waking experience.

Rats were trained on a spatial memory task, and hippocampal and cortical activity was recorded during both behavior and subsequent memory consolidation during the first 2 hours

other spikes





Fig. 3. Delta waves isolate cortical computations. (**A**) Cross-correlations (curves and shaded areas, mean \pm SEM) between hippocampal ripple activity (fixed, 0 s) and prefrontal activity (sliding window). Observed cross-correlations (orange) were significantly different from a time-shifted control (gray) for cortical activity after ripples (horizontal orange line: Monte Carlo test, *P* < 0.05). Delta waves (dashed line; peak occurrence rate 130 ms after ripples) tend to occur within the critical window when prefrontal activity remains correlated with the preceding ripple activity. (**B**) Simultaneous recording of prefrontal and hippocampal activity around a delta wave (gray-shaded rectangle). (Top) Proportion of prefrontal spikes predicted by the firing of hippocampal cells (partner spikes). (Center) Raster plot of spikes

emitted by 68 simultaneously recorded prefrontal units (red ticks: partner spikes; gray ticks: other spikes). (Bottom) Simultaneously recorded LFPs in the mPFC (blue: delta wave) and hippocampus (broadband and ripple-band filtered signal; blue: ripples). During delta waves, partner spikes occurred in isolation (red ellipse). Partner spikes emitted by the same units outside delta waves (gray ellipses) formed a considerably smaller proportion of the ongoing cortical activity. Black calibration bar: 0.5 s. Filt, filtered. (**C**) Signal-to-noise ratio (curves and shaded areas, mean \pm SEM) of partner spikes relative to other spikes around delta waves. Observed values (blue) were significantly different from a time-shifted control (gray) during delta waves (horizontal blue line: Monte Carlo test, P < 0.05).

of sleep (5). Hippocampal spiking activity during ripples was significantly correlated with cortical delta spikes that occurred immediately (50 to 200 ms) afterward (the effect was not due to data recorded from any single

rat) (Fig. 2, A and B, and fig. S8). This increased correlation was due to a large proportion of positively correlated interregional pairs of neurons (Fig. 2C). Furthermore, ripple spikes were better correlated with delta spikes than with spikes occurring at similar delays during an up state (correlations were greater in the presence of a delta wave) (fig. S9). A generalized linear model (GLM) analysis showed that ensemble activity in the hippocampus





Fig. 4. Induction of memory consolidation by isolation of partner spikes. (A) (Left) Experimental protocol. Delta waves were induced by brief single-pulse electrical stimulation of deep cortical layers. Induced delta waves were triggered to isolate either partner activity (coupled stimulation: green, 130 ms after ripples) or other cortical activity (delayed stimulation; purple, 290 to 370 ms after ripples) during sleep after limited training on a spatial object-recognition task. (Right) Object discrimination index during the recall phase. Only delta waves triggered to isolate partner activity (coupled stimulation) resulted in memory consolidation and enhanced task performance. (B) Performance of a GLM trained to predict delta spikes on the basis of preceding hippocampal activity (200-ms window), measured as

percent improvement relative to a shuffled control (prediction gain). Only delta waves triggered to isolate partner activity resulted in a significant prediction of delta spikes (P = 0.0030 for isolation of partner spikes by coupled stimulation: P = 0.1301 for isolation of other spikes by delayed stimulation: Wilcoxon rank sum test). stim, stimulation. **P < 0.01. (C) Cross-correlation (curves and shaded areas, mean ± SEM) of hippocampal activity and delta spikes (green: isolation of partner spikes by coupled stimulation; purple: isolation of other spikes by delayed stimulation; gray: time-shifted control; horizontal green line: Monte Carlo test, P < 0.05). (**D**) Enrichment in positive correlations upon isolation of partner spikes by coupled stimulation (top) but not of other spikes by delayed stimulation (bottom).

could significantly predict which cortical cells would emit delta spikes (Fig. 2D). In contrast, delta spikes could not be predicted from the combined activity of all hippocampal units that ignored cell identity (multiunit activity), ruling out the possibility that delta spikes merely reflect the overall level of hippocampal excitatory drive during ripples. Finally, ripples facilitated (but did not entirely control) the formation of delta assemblies (fig. S7). Furthermore, the same GLM analysis applied to hippocampal and cortical ensembles showed that hippocampal activity could even predict delta components (Fig. 2D).

Our second prediction concerned the relation of this predictive bias to behavior. In sleep sessions preceding the task, hippocampal spiking activity during ripples failed to predict subsequent delta spikes or assemblies (fig. S10), indicating that the predictive bias emerged after task performance. We then investigated the behavioral correlates of the prefrontal units whose delta spikes were significantly predicted by hippocampal ripple activity during sleep after behavior ("partner cells") (fig. S11 and tables S1 and S2). These cortical cells displayed higher levels of task-relevant firing during behavior (Fig. 2E) (we failed to find a similar effect for delta components, possibly because of low statistical power due to their limited number: n = 14 components, n = 9predicted). We further investigated the behavioral correlates of delta assemblies and found that these assemblies were also expressed during task performance but not outside delta waves nor in sleep that preceded behavior (fig. S12).

These observations suggest that in addition to triggering the reorganization of cortical subnetworks during the transition to the up state (5), an unsuspected role of the delta wave may be to isolate from interference specific cortical computations taking place in response to hippocampal replay. Consistent with this idea, delta waves typically occurred within a critical time window when cortical activity remained correlated with hippocampal ripple activity (Fig. 3A). To test whether delta waves tended to preferentially silence cortical activity that was related to the ongoing hippocampo-cortical dialogue, we classified individual prefrontal spikes as "partner spikes" if they followed spikes emitted by their significantly correlated hippocampal units or "other spikes" if they were unrelated to the preceding hippocampal activity (Fig. 3B) (more examples are shown in fig. S13). The signal-to-noise ratio for partner spikes peaked during delta waves (Fig. 3C), and this was due to the selective silencing of nonpartner activity during delta waves (fig. S14).

Does this isolation of cortical computations play a critical role in memory consolidation? A prediction of this hypothesis is that isolating cortical assemblies by experimental induction of delta waves should trigger memory consolidation, but only if the isolated activity is relevant to the hippocampo-cortical dialogue (partner spikes). We have already shown that triggering delta waves when endogenous mechanisms fail to do so can boost memory consolidation, provided that the delta waves are induced in an appropriate time window (Fig. 4A) (5). We thus sought to confirm the prediction that these delta waves actually isolated partner spikes (that delta spikes did occur during induced delta waves and that they were predicted by hippocampal activity). Similar to our observations in natural sleep (above), stimulation-induced delta waves did feature spiking activity (fig. S15), and these delta spikes were predicted by preceding hippocampal activity coinciding with the timing of ripples (Fig. 4, B to D). In contrast, slightly delaying the induction of delta waves (by ~200 ms) (Fig. 3A) to isolate nonpartner delta spikes (Fig. 4, B to D) failed to induce memory consolidation (5).

Our results challenge the generally accepted tenet that delta waves, reflecting the down states of the sleep slow oscillation, are periods of complete cortical silence (1, 6, 25), to the point that they have sometimes been defined as such (26, 27) and that occasional spikes have been routinely ignored when detected (28, 29). We focused on delta spikes and found that they are not neuronal noise due to imperfect silencing of the cortical mantle. On the contrary, they constitute a common phenomenon potentially implicating all neurons and all delta waves, and they reflect genuine processing involved in memory consolidation.

This observation also provides a mechanism for the documented but puzzling role of delta waves in memory consolidation: Synchronized silence across most of the cortex isolates the network from competing inputs while a select subpopulation of neurons maintains relevant spike patterns between epochs of hippocampo-cortical information transfer (10, 12, 14) and epochs of cortical plasticity (15, 16) and network reorganization (5, 18, 19). Yet, in many cases, cortical activity during delta waves could not be reliably predicted from the preceding hippocampal ripple activity. Such cortical activity could instead have been related to interactions with other brain networks. This suggests that delta spikes and assemblies might constitute a general mechanism of isolated cortical computation beyond the hippocampo-cortical dialogue.

REFERENCES AND NOTES

- 1. M. Steriade, A. Nuñez, F. Amzica, J. Neurosci, 13, 3252-3265 (1993)
- 2 H.-V. V. Ngo, T. Martinetz, J. Born, M. Mölle, Neuron 78. 545-553 (2013).
- 3 L. Marshall, H. Helgadóttir, M. Mölle, J. Born, Nature 444, 610-613 (2006).
- S. Chauvette, J. Seigneur, I. Timofeev, Neuron 75, 1105-1113 (2012).
- N. Maingret, G. Girardeau, R. Todorova, M. Goutierre, M. Zugaro, Nat. Neurosci. 19, 959-964 (2016).
- 6. A. Sirota, G. Buzsáki, Thalamus Relat. Syst. 3, 245-259 (2005).
- A. Sirota, J. Csicsvari, D. Buhl, G. Buzsáki, Proc. Natl. Acad. Sci. U.S.A. 100, 2065-2069 (2003).
- 8. G. Girardeau, K. Benchenane, S. I. Wiener, G. Buzsáki,
- M. B. Zugaro, Nat. Neurosci, 12, 1222-1223 (2009) V. Ego-Stengel, M. A. Wilson, Hippocampus 20, 1-10 (2010).
- 10. A. K. Lee, M. A. Wilson, Neuron 36, 1183-1194 (2002).
- 11. D. Ji, M. A. Wilson, Nat. Neurosci. 10, 100-107 (2007).
- 12. G. Rothschild, E. Eban, L. M. Frank, Nat. Neurosci. 20, 251-259 (2017).
- 13. C. M. Wierzynski, E. V. Lubenov, M. Gu, A. G. Siapas, Neuron 61. 587-596 (2009).
- 14. A. Peyrache, M. Khamassi, K. Benchenane, S. I. Wiener,
- F. P. Battaglia, Nat. Neurosci. 12, 919-926 (2009).
- 15. P. B. Kruskal, L. Li, J. N. MacLean, Nat. Commun. 4, 2574 (2013)
- 16. T. Gulati, L. Guo, D. S. Ramanathan, A. Bodepudi, K. Ganguly, Nat. Neurosci. 20, 1277-1284 (2017).
- 17. A. G. Siapas, M. A. Wilson, Neuron 21, 1123-1128 (1998). 18. M. Rosanova, D. Ulrich, J. Neurosci. 25, 9398-9405 (2005).
- 19. J. Seibt et al., Nat. Commun. 8, 684 (2017).
- 20. D. Marr, Philos. Trans. R. Soc. London Ser. B 262, 23-81 (1971).
- 21. G. Buzsáki, Neuroscience 31, 551-570 (1989).
- 22. P. W. Frankland, B. Bontempi, Nat. Rev. Neurosci. 6, 119-130 (2005).
- 23. V. Lopes-dos Santos, S. Ribeiro, A. B. L. Tort, J. Neurosci. Methods 220, 149-166 (2013).
- 24. K. D. Harris, J. Csicsvari, H. Hirase, G. Dragoi, G. Buzsáki, Nature 424, 552-556 (2003).
- 25. M. Steriade, I. Timofeev, Neuron 37, 563-576 (2003).
- 26. A. Luczak, P. Barthó, S. L. Marguet, G. Buzsáki, K. D. Harris, Proc. Natl. Acad. Sci. U.S.A. 104, 347-352 (2007).
- 27. L. A. Johnson, D. R. Euston, M. Tatsuno, B. L. McNaughton, J. Neurosci. 30, 2650-2661 (2010).
- 28. J. Csicsvari et al., J. Neurophysiol. 90, 1314-1323 (2003). 29. D. Jercog et al., eLife 6, e22425 (2017).
- 30. N. Maingret, R. Todorova, M. Zugaro, Recordings and timed stimulation of rat dorsal hippocampal area CA1 and medial prefrontal cortex during behavior and sleep. CRCNS (2019): https://doi.org/10.6080/K0TD9VJG.

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SUPPLEMENTARY MATERIALS

science.sciencemag.org/content/366/6463/377/suppl/DC1 Materials and Methods Figs. S1 to S15 Tables S1 and S2 References (31-34)

View/request a protocol for this paper from Bio-protocol.

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mechanisms for O=O bond formation. In 2016, a study reported a room-temperature structure of PSII following two light flashes, hence enriched in the $\mathbf{S}_{_{\!3}}$ state of the OEC (11). It reported no new S₂-state electron density around Mn1, as one would expect for a newly bound water. This result directly conflicts with the proposed Siegbahn model and EPR studies. Shortly following this study, another group reported an S₂ structure with new electron density near Mn1 and O5 that was assigned to a newly bound oxygen (O6) (12). This was considered support for the S_ainsertion substrate water model. However, the study showed close proximity (0.15 nm) between O5 and O6, consistent with O-O bond formation producing a peroxide at the S_a state, which would in turn be associated with Mn reduction in the S_a-to-S_a transition. This is in conflict with EPR and x-ray spectroscopic data (3, 9, 14, 15). A more recent study reports a new oxygen "Ox" bound both to Mn1 and the Ca ion of the OEC (13), and with a 0.21 nm distance to O5, much longer than the reported 0.15 nm (12) and too long to represent the O-O bond of a peroxide.

Suga et al. have now cryotrapped serially flashed microcrystals of PSII with the goal of achieving more accurate interatomic distances, especially between O5 and O6. In the S_o state, they observed an open cubane structure with a five-coordinate Mn1. At the S_a state, a flip in the side chain of the monodentate carboxylate ligand Glu189 (12) provides room for O6 to insert and bind to Mn1 and Ca. To address the crucial O5-O6 distance, Suga et al. calculated difference maps as a function of modeled O5-O6 distance and found the smallest residual density at a distance of about 0.19 nm. Their full residual density analysis suggests an oxyl/oxo pairing for O5 and O6, which they consider evidence for an oxyl/oxo coupling mechanism in the O=O bond formation at the final S_4 state. Glu¹⁸⁹ appears to gate water insertion into the OEC from the "O1" water channel of the PSII reaction center.

Although there are still open questions as to the detailed assignments of substrate waters and the exact modes of water oxidation and O=O bond formation, the results from different studies are starting to converge, and the structural results appear to be closely in line with mechanistic proposals supported by computational chemistry and spectroscopy. The field anxiously awaits possible timeresolved structures detailing the O=O bond formation through the S_4 -to- S_0 transition.

REFERENCES AND NOTES

- M. Suga et al., Science 366, 334 (2019) 1. 2.
- D. J. Vinyard, G. W. Brudvig, Annu. Rev. Phys. Chem. 68, 101 (2017).
- D.A. Pantazis, ACS Catal. 8, 9477 (2018). 3 4
- A. Zouni et al., Nature 409, 739 (2001) K.N. Ferreira et al., Science 303, 1831 (2004). 5
- Y. Umena et al., Nature 473, 55 (2011). 6.
- J. M. Peloquin et al., J. Am. Chem. Soc. 122, 10926 (2000). 7
- 8 P.E.M. Siegbahn, Biochim. Biophys. Acta Bioenerg. 1827, 1003(2013)
- 9 N. Cox et al., Science 345, 804 (2014).
- 10. M. Chrysina et al., Proc. Natl. Acad. Sci. U.S.A. 116, 16841 (2019)
- 11 I. D. Young et al., Nature 540, 453 (2016).
- M. Suga et al., Nature 543, 131 (2017). 12
- 13 J. Kern et al., Nature 563, 421 (2018)
- 14 M. Haumann et al., Science 310, 1019 (2005)
- 15. N. Schuth et al., Inorg. Chem. 57, 10424 (2018).

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The S states in the oxygen-evolution reaction

The oxygen-evolving complex is photo-oxidized through a series of S states to produce molecular oxygen from water. In the final steps before O=O bond formation, a new oxygen, O6, binds to the vacant site at Mn1. After a final photo-oxidation event, 05 and 06 appear poised to form an 0=0 bond, releasing molecular oxygen, reducing the cluster, and beginning the catalytic cycle anew. Glutamatic acid at position 189 is noted as E189.



NEUROSCIENCE

Spikes in the sleeping brain

Memory is replayed and consolidated under low background noise during deep sleep

By Yuji Ikegaya^{1,2} and Nobuyoshi Matsumoto¹

emories of recent experiences are transferred and fixed in long-term storage in the neocortex during sleep. A key aspect of this process is the occurrence of ripples, which are high-frequency oscillations in neuronal activity in the hippocampus (1, 2). Ensembles of hippocampal neurons that have been activated during an experience emit synchronized neuronal activity (spikes) during ripples in subsequent sleep (3). These "replayed" memories propagate to the neocortex during slow-wave states (4), which are characterized by alternating "bright" states with active neurons and "dark" states, called delta waves, without active neurons (5). Researchers reasoned that hippocampal-cortical dialogue occurs during the bright states and that delta waves represent intermittent neocortical "sleep" to recover from synaptic fatigue (6) or increased potassium conductance (7) and have no active function. On page 377 of this issue, Todorova and Zugaro (8) challenge this view, demonstrating that a small number of neurons are reactivated during delta waves.

Todorova and Zugaro recorded activity, both spikes from multiple neurons and local field potentials, simultaneously in the rat dorsal hippocampus and the medial prefrontal cortex, which is believed to store long-term memories. They reconfirmed that the prefrontal cortex becomes inactive during each delta-wave epoch but also discovered that a few neurons remain active and occasionally emit synchronous spikes during delta waves (which they called "delta spikes") (see the figure). The neurons recorded in this study constitute only a small portion of the total cells in the prefrontal cortex. Thus, it is important to ask how frequently delta spikes occur. The local field

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GRAPHIC: A. KITTERMAN/SCIENCE

Hippocampalcortical dialogues during sleep

The hippocampus communicates with the neocortex and activates experience-associated neocortical neuron ensembles during delta waves, low-cerebral activity states that emerge intermittently during slow-wave sleep. This selective activity focuses memory consolidation.



potential waveforms, which reflect the synchronous activity of a group of neurons, do not differ depending on whether delta waves are accompanied by delta spikes in the recorded neurons. Moreover, the probability of detecting delta spikes is proportional to the number of simultaneously recorded neurons. Extrapolating these observations suggests that delta spikes accompany virtually all delta waves.

Delta spikes are not shots in the dark or remnants of ripples that accidentally fail to disappear during delta waves. Rather, delta spikes occur in spatiotemporally designed, specifically recursive configurations and seem to be actively generated in the brain.

Todorova and Zugaro annotated delta spikes as behavioral correlates. As rats were trained in a spatial memory task, behaviorally relevant neurons became preferentially activated in delta waves during the following sleep period. The neuronal replay in the prefrontal cortex appears to be

instructed by hippocampal ripples, because the set of prefrontal neurons that emit a delta spike in a given delta wave can be statistically predicted from the set of hippocampal neurons activated in the ripple immediately preceding the delta wave. The spike correlations of cell pairs between the hippocampus and the medial prefrontal cortex remain high for hundreds of milliseconds, indicating that hippocampal-cortical dialogue occurs within a short time frame, during which the neocortex serves as a temporary reservoir of hippocampal information. As a result, memory-relevant delta spikes are fired mainly in the early phase of each delta-wave event.

Delta spikes consist mainly of behaviorally relevant neurons, whereas behaviorally irrelevant neurons are more likely silenced during delta waves. Reducing irrelevant information improves the signal-to-noise ratio of neuronal information and thereby helps isolate cortical computations to avoid deleterious cross-talk. That is, the neocortex quiets down during delta waves and focuses attention on the content to be memorized, consistent with the authors' previous prediction that delta waves "isolate target synapses from competing inputs, allowing selective reorganization of the network" (9). This leads to the possibility that facilitating delta spikes in ripple-associated neurons could enhance memory consolidation. Consistent with this idea, task performance in mice is enhanced when artificial delta waves containing ripple-coupled delta spikes are

"...task performance in mice is enhanced when artificial delta waves...are repeatedly induced..."

le-coupled delta spikes are repeatedly induced using brief single-pulse electrical stimulation of deep cortical layers during sleep after an object-discrimination task. This suggests that delta spikes mediate memory consolidation.

How do delta spikes contribute to memory consolidation? It is unknown

what delta waves give rise to in prefrontal neuronal circuits. Delta spikes are transient and thus are unlikely to work as persistent memories by themselves. So, what form of long-lasting plasticity is induced by delta spikes? Moreover, activity patterns in individual task-associated neocortical neurons change over a period of days after learning (10). Do delta spikes also have an instructive role in the dynamic reorganization of neuronal identities? Another line of evidence suggests that delta waves are involved in forgetting rather than memory consolidation (11). Thus, drifting delta spikes could contribute to deactivation of memory traces.

Another question is how the dorsal hippocampus and the medial prefrontal cortex communicate during slow-wave sleep. Direct synaptic connections are unlikely to exist between these brain regions. On the basis of anatomical evidence of axonal projections, candidates for the relay station include the ventral hippocampus (12), a neocortical area via the entorhinal cortex (13), and the retrosplenial cortex via the subiculum (14). Recording or ablation of neuronal activity in these brain regions is necessary to answer this question.

Delta waves are a hallmark of the entire neocortex during sleep, and delta spikes may occur in cortical regions other than the medial prefrontal cortex. Do delta spikes have a role in processes other than memory consolidation? Todorova and Zugaro have successfully linked their unexpected discovery to the current knowledge of memory and sleep, but future investigations are still required to uncover the precise mechanisms and functions of delta spikes.

REFERENCES AND NOTES

- G. Girardeau, K. Benchenane, S. I. Wiener, G. Buzsáki, M. B. Zugaro, *Nat. Neurosci.* 12, 1222 (2009).
- 2. A. Fernández-Ruiz et al., Science 364, 1082 (2019).
- 3. G. Buzsáki, *Neuroscience* **31**, 551 (1989).
- 4. N.K. Logothetis et al., Nature 491, 547 (2012).
- 5. F. Siclari, G. Tononi, *Curr. Opin. Neurobiol.* **44**, 222 (2017).
- 6. D. Contreras, I. Timofeev, M. Steriade, *J. Physiol.* **494**, 251 (1996).
- M. V. Sanchez-Vives, D. A. McCormick, *Nat. Neurosci.* 3, 1027 (2000).
- 8. R. Todorova, M. Zugaro, *Science* **366**, 377 (2019).
- N. Maingret, G. Girardeau, R. Todorova, M. Goutierre, M. Zugaro, Nat. Neurosci. 19, 959 (2016).
- L. N. Driscoll, N. L. Pettit, M. Minderer, S. N. Chettih, C. D. Harvey, *Cell* **170**, 986 (2017).
- 11. J. Kim, T. Gulati, K. Ganguly, Cell **179**, 514 (2019).
- A. M. Thierry, Y. Gioanni, E. Dégénétais, J. Glowinski, Hippocampus 10, 411 (2000).
- 13. R. Insausti, M. T. Herrero, M. P. Witter, *Hippocampus* 7, 146 (1997).
- M. P. Witter, R. H. Ostendorf, H. J. Groenewegen, *Eur. J. Neurosci.* 2, 718 (1990).

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Materials and Methods

Animals

The analyses presented here were performed on data collected for a previous study by Maingret *et al.* (5). Briefly, a total of 9 male Long Evans rats (René Janvier, Le Genest, St Isle, France; weight, 280–350 g) were maintained on a 12h:12h light-dark cycle (lights on at 7 am). Training and experiments took place during the day. Rats were grouphoused until one week before surgery. All experiments were in accord with institutional (CNRS Comité Opérationnel pour l'Éthique dans les Sciences de la Vie) and international (US National Institutes of Health guidelines) standards, legal regulations (Certificat no. B751756), and ethical requirements (Ethics Committee approval #2012-0048) regarding the use and care of animals.

Surgery

Rats (n = 9) were deeply anesthetized (xylazine, 0.1 ml intramuscular; pentobarbital, 40 mg per kg of body weight, intraperitoneal; 0.1 ml pentobarbital supplemented every hour) and implanted with a custom-built microdrive with 16 individual tetrodes (groups of four twisted 12 µm tungsten wires, gold-plated to ~200 kΩ), of which 8 targeted the prelimbic and infralimbic regions of the right mPFC (AP: +2.7 mm from bregma; ML: +1.5 mm, angled at 10° from the sagittal plane), and the other 8 targeted the CA1 subfield of the right hippocampus (AP: -3.5 to -5.5 mm; ML: +2.5 to +5.0 mm). A custombuilt bipolar stimulation electrode consisting of two stainless steel wires (total length, 1.5 mm; inter-wire interval, 0.5 mm; wire diameter, 70 µm) was implanted in the left neocortex (AP: +2 mm; ML: -2 mm; DV: -1.5 mm from the *dura*, motor area). Miniature stainless steel screws (reference and ground) were implanted above the cerebellum. During recovery from surgery (minimum 3 days), the rats received food and water ad libitum. The recording electrodes were then progressively lowered until they reached their targets and then adjusted every day to optimize yield and stability.

Behavioral task

The spatial object recognition task has been described previously (5). Briefly, during a 3 min encoding phase, the rats explored a $70 \text{ cm} \times 50 \text{ cm}$ arena with two identical objects placed in two adjacent corners. The rats were then placed in a flower pot for sleep sessions, which lasted until 1,000 stimulations had been delivered (see below; ~4,000 s of SWS), then returned to their home cage. On the following day, during a 5 min recall phase, the rats explored the same arena, in which one of the objects had been displaced to the opposite corner. The rats were then placed in a flower pot for uninterrupted sleep sessions without stimulation. After an interval of at least two days, each rat performed the task a second time with different objects. The order of the stimulation conditions was pseudo-randomly distributed among rats.

Stimulation protocol

The stimulation protocol has been described previously (5). Briefly, threshold crossing on the ripple band-filtered hippocampal signal automatically triggered a monophasic singlepulse (0.1 ms) stimulation of the deep layers of the motor cortex, delivered by a constant current stimulator (SD9 square pulse stimulator, Grass Technologies). For each animal, the minimum voltage necessary to reliably induce propagating delta waves was determined prior to training (range: 17.5-22.5 V). The number of stimulations was limited to one every two seconds, and the total number of stimulations was set to 1,000, yielding a stimulation period of ~4,000 s. To isolate partner cells, stimulations were delivered to trigger a delta wave ~130 ms following SPW-R detection, emulating endogenous fine-tuned SPW-R-delta coordination. To isolate other (non-partner) cells, an additional random delay (range: 160-240 ms) was introduced between SPW-R detection and stimulation onset.

Data acquisition and processing

Brain signals were preamplified (unity-gain headstages, Noted Bt, Pécs, Hungary), amplified $\times 500$ (Neuralynx L8, Bozeman, MT, USA), acquired and digitized with two synchronized Power1401 systems (CED, Cambridge, UK). A red LED was used to track the

instantaneous position of the animals (25 Hz). For off-line spike sorting, the wide-band signals were converted, digitally high-pass filtered (nonlinear median-based filter) and thresholded, and waveforms were extracted and projected to a PCA subspace using NDManager (L. Hazan and M. Zugaro, http://neurosuite.sourceforge.net, (31)). Spike sorting used a semi-automatic cluster cutting procedure combining KlustaKwik (K.D. Harris, http://klustakwik.sourceforge.net) and Klusters (L. Hazan, http://neurosuite. sourceforge.net, (31)). In the prefrontal cortex, excitatory and inhibitory cells were discriminated based on significant short-latency peaks and troughs in cross-correlograms (32), that were further validated by visual inspection. In addition, units were divided into putative pyramidal cells and interneurons based on half-amplitude duration and trough to peak time (32). In the hippocampus, putative pyramidal cells were identified based on firing rate and bursting properties. Neurophysiological and behavioral data were explored using NeuroScope (L. Hazan, http://neurosuite.sourceforge.net, (31)). LFPs were derived from wideband signals by downsampling all channels to 1,250 Hz.

At the end of the experiments, recording sites were marked with small electrolytic lesions. Rats were deeply anesthetized with a lethal dose of pentobarbital, and intracardially perfused with saline (0.9%) followed by paraformaldehyde (10%). Coronal slices $(40 \,\mu\text{m})$ were stained with cresyl-violet.

Data analysis and statistics

Data were analyzed using in Matlab (MathWorks, Natick, MA), using FMAToolbox (http://fmatoolbox.sourceforge.net) and custom written programs. Spectrograms were constructed using Chronux (http://chronux.org/).

Object responsivity

Object responsivity was computed as described previously (5). Briefly, the responsivity index R was defined as the mean firing rate r over the quadrants containing the objects, and z-scored relative to the distribution F of firing rates in the empty quadrants, i.e. $R = (r - \mu)/\sigma$ where μ and σ are the mean and standard deviation of F. Thus, the object responsivity index R measured by how much, relative to its baseline variability, a cell increased its firing rate around the objects.

Sleep scoring

Sleep stages (SWS vs REM) were determined by automatic K-means clustering of the theta/delta ratio extracted from the power spectrograms during the episodes where the animal was immobile (linear velocity <3 cm/s for at least 30 s, with brief movements <0.5 s).

Ripple detection

To detect ripple events, we first detrended the LFP signals and used the Hilbert transform to compute the ripple band (100–250 Hz) amplitude for each channel recorded from the CA1 pyramidal layer. We then averaged these amplitudes, yielding the mean instantaneous ripple amplitude. To exclude events of high spectral power not specific to the ripple band, we then subtracted the mean high-frequency (300–500 Hz) amplitude (if the difference was negative, we set it to 0). Finally, we z-scored this signal, yielding a corrected and normalized ripple amplitude R(t). Ripples were defined as events where R(t) crossed a threshold of 3 s.d. and remained above 1 s.d. for 30 to 110 ms.

Delta wave detection

Delta waves were identified based on detection of both large positive deflections in the LFP and concurrent decreases in multiunit activity. First, the LFP recorded from each tetrode located in the mPFC was filtered (0–6 Hz) and z-scored, yielding D(t). The beginning $(t_{beginning})$, peak (t_{peak}) , and end (t_{end}) of putative delta waves were defined as upward-downward-upward zero-crossings of D'(t). Epochs where either $D(t_{peak}) > 2$ and $D(t_{end}) < 0$, or $D(t_{peak}) > 1$ and $D(t_{end}) < -1.5$, were deemed candidate events. Candidate events briefer than 150 ms or longer than 500 ms were discarded. Second, the instantaneous mPFC multiunit activity was smoothed (Gaussian window, $\sigma = 60$ ms). Candidate events where the smoothed activity decreased relative to a 2s period around t_{peak} were considered delta waves.

Delta spikes and assemblies

To quantify spiking during delta waves, we computed the distribution of delays between a delta wave peak and the nearest spike emitted by each neuron recorded on the same tetrode as the delta wave. This yielded a bimodal distribution (Fig. 1C): while most nearest spikes occurred 100–200 ms from the delta wave peak (i.e. in the UP state), in a minority of cases a spike occurred within 15 ms of the delta wave peak. Using this value as a conservative threshold, we defined delta spikes as spikes emitted within 15 ms of the peak of a delta wave detected on the same tetrode as the active neuron (all analyses were repeated and confirmed using ± 30 ms time windows). Delta waves in which we did not detect any delta spikes were referred to as silent delta waves.

We used independent component analysis (ICA) to detect significant cell assemblies (33). We computed a delta spike matrix M, where M(i, j) is the number of spikes emitted by neuron i within 15 ms of delta wave j. We z-scored M and projected it onto its n significant principal components – the n components with eigenvalues exceeding the Marcenko-Pastur threshold λ_{max} . We then ran ICA (using the FastICA package for Matlab, http://research.ics.aalto.fi/ica/fastica) on the projected matrix to obtain the weight vectors describing the assemblies. The activation of a given cell assembly k was computed as $A_k = M^T P_k M$, where P_k is the projection operator of the cell assembly (the outer product of its weight vector) with its diagonal set to zero.

To assess whether the detected activity patterns were also expressed in other behavioral conditions or brain states (Fig. S12), we detected cell assemblies separately in five different conditions: delta waves in pre-task and post-task sleep, non-delta periods in pre-task and post-task sleep (100 ms non-overlapping bins excluding delta waves), and task-related behavior (100 ms non-overlapping bins). To estimate similarity across the patterns in any two conditions, each component in one condition was matched to the component in the other condition which yielded the highest absolute correlation, and the average correlation between matched components was computed (adapted from van de Ven *et al.* (33)). To assess significance, we compared this average correlation to a distribution of

shuffled correlations obtained by permuting the weights of each component across cells and reiterating the above procedure (matching and computing average correlation between matched components) 1,000 times.

Cooperative activity within delta waves was assessed by quantifying to what extent the delta spike activity of each prefrontal cortical cell could be predicted from the delta spike activity of all other prefrontal cortical cells ('peer prediction' (24)). Thus, for each prefrontal cortical neuron *i*, delta waves were split into n_i non-overlapping partitions, ensuring that each partition contained exactly one delta wave where neuron *i* fired. Using a subset of $(n_i - 1)$ partitions, a Generalized Linear Model (GLM) was trained to predict the binary delta spike vector D_i of neuron *i*, from the matrix $D_{(i)}$ containing the delta spike counts of all other prefrontal neurons (i.e. excluding neuron *i*). The GLM was tested on the remaining partition. This procedure was repeated n_i times, each time using a different subset of training and test partitions, resulting in multi-fold cross-validation. Following (12), the quality of the prediction was assessed by comparing the median prediction error *e* to the median error $e_{shuffled}$ obtained by shuffling 1,000 times the predictions relative to the observed data D_i . The prediction gain *g* was defined as $g = e_{shuffled}/e$.

Prediction of delta spikes by hippocampal activity

Cross-correlations. Cross-correlations between hippocampal ripple activity and surrounding prefrontal cortical activity (shown in Fig. 3A) were computed as follows. For each hippocampal pyramidal unit *i*, spikes were counted in 200 ms windows centered on each t_{ripple} , yielding a spike count vector N_i . Similarly, for each prefrontal cortical unit *j*, spikes were counted in 200 ms windows centered on $t_{ripple} + k$ ($k \in [-500, 500]$ ms), yielding a spike count vector $N_j(k)$ for each temporal shift *k*. The Spearman rank-order correlation $\rho_{ij}(k)$ between N_i and $N_j(k)$ was averaged over *i* and *j*, yielding the mean cross-correlation $\rho(k)$ as a function of the temporal shift *k*.

Cross-correlations between delta spikes and surrounding hippocampal ripple activity (shown in Fig. 2B) were computed using a similar procedure, where delta spikes were counted in 30 ms windows centered on t_{delta} (corresponding to the 15 ms threshold dis-

tance defined in Fig. 1C), and hippocampal ripple spikes were counted in 200 ms windows centered on $t_{delta} + k$. Two additional criteria were applied for inclusion in this analysis. Firstly, for a given prefrontal cortical unit *i*, only delta waves recorded from the same tetrode were considered. Secondly, for a given value of *k*, delta waves were taken into account only when ripple activity took place within a 200 ms window centered on $t_{delta} + k$. Prefrontal cortical cells that participated in at least one significant cross-structural cell pair for k = -200 ms were defined as partner cells.

Cross-correlations between hippocampal spikes and delta assemblies were computed using the same procedure, but using assembly activations A_i (see section *Delta spikes and assemblies*, above), rather than spike counts, during delta waves.

To assess the significance of the cross-correlations described above, we generated control cross-correlations $\rho_{ij}^{shifted}(k)$ by shifting one of the temporal series: for instance, hippocampal activity surrounding the *n*-th delta wave was correlated with delta spikes emitted during the (n+1)-th delta wave, thus shifting the correlation by one delta wave. This approach, as opposed to a completely random shuffle of all events, selectively disrupted fine timescale correlations but preserved long timescale trends, effectively preventing false positives due to non-specific factors such as sleep progression or depth.

When these analyses were repeated after removing from the spike trains all hippocampal spikes emitted outside ripples, all results were confirmed.

Enrichment (Figs. 2C and 4D) was defined as the difference between observed and spurious correlations $\rho_{ij}(k) - \rho_{ij}^{shifted}(k)$.

Generalized Linear Model (GLM). The predictive power of hippocampal ripple activity on prefrontal delta spikes was further assessed using a GLM analysis, which considered ensemble activity rather than neuron pairs (as in the cross-correlation analyses above). The procedure was similar to the peer prediction method described for prefrontal cooperative activity during delta waves. Briefly, a GLM was trained to predict the binary delta spike vector D_i of prefrontal cortical neuron i, from the matrix H containing the spike counts of hippocampal pyramidal neurons in 200 ms windows preceding delta waves $(H_{mj}$ is the number of spikes emitted by neuron j before the m-th delta wave). The same multi-fold cross-validation scheme was used as above, yielding a prediction gain $g = e_{shuffled}/e$. When these analyses were repeated after removing from the spike trains all hippocampal spikes emitted outside ripples, all results were confirmed.

Predicted spikes and signal-to-noise ratio. This analysis examined hippocampocortical correlations around each delta wave k (not restricted to delta spikes). As in the previous cross-correlation analyses, partner cells were first identified based on significant Spearman's rank correlation (here, between prefrontal $N_i(k)$ and hippocampal $N_j(k - 200 \text{ ms})$). Subsequently, for each prefrontal cortical unit, individual spikes were deemed 'partner spikes' if any of the unit's partner cells in the hippocampus fired within a preceding 200 ms window. The signal-to-noise ratio measures the ratio of partner spikes versus other spikes in 20 ms bins around delta waves.



Fig. S1. Delta spike waveforms. (A) Spike waveforms for all the recorded delta spikes (black traces) for two example units. Colored traces: average waveforms for all spikes that the same units fired outside delta waves. (B) L-ratio was not different (P = 0.72279, Wilcoxon rank-sum test) between delta spikes and a random sample of non-delta spikes. (C) Isolation distance was not different (P = 0.84504, Wilcoxon rank-sum test) between delta spikes and a random sample of non-delta spikes and a random sample of non-delta spikes.



Fig. S2. Delta wave firing across units. (A) During SWS, the proportion of spikes fired in delta waves did not differ between CCG-identified inhibitory (n = 17), excitatory (n = 57) and undefined (n = 366) cells (P = 0.5322), Kruskal-Wallis test). Inset: units were additionally divided according to waveform. The proportion of spikes fired in delta waves did not differ between waveform-identified putative interneurons and putative pyramidal cells (P = 0.0734), Wilcoxon rank-sum test). **(B)** There was no correlation (P > 0.05) between firing rate and the proportion of SWS spikes fired in delta waves for any of the cell types (top, excitatory cells; middle, inhibitory cells; bottom: undefined cells).



Fig. S3. Timing of silent and non-silent delta waves. (A) Delta wave duration was not different between silent (n = 21, 733) and non-silent (n = 7, 162) delta waves (P = 0.45582, Wilcoxon rank-sum test). **(B)** The proportion of non-silent delta waves did not vary across sleep (P > 0.05, binomial proportion tests). **(C)** The proportion of SWS spikes fired in delta waves for each unit was not different between pre-task (blue) and post-task (green) sleep (P = 0.78105, Wilcoxon signed rank test). **(D)** Delta wave timing with respect to other delta waves was not different between silent and non-silent delta waves. Left: delta-wave occurrence rate centered on silent (black) and non-silent (red) delta waves (curves and shaded area, mean \pm s.e.m.) Note that silent delta waves. Center: median inter-delta intervals were not significantly different following silent vs non-silent delta waves (P = 0.76086, Wilcoxon signed rank test; bars: median \pm s.e.median; dots: within-session medians). Right: median inter-delta intervals were not significantly different significantly different preceding silent vs non-silent delta waves (P = 0.80774, Wilcoxon signed rank test; bars: median \pm s.e.median; dots: within-session medians).



Fig. S4. Delta spikes recorded from different cortical layers. (A) Top: example histological sections showing electrode penetrations at various depths. Bottom: average waveforms for delta waves recorded from example electrodes in each rat (red: putative more superficial recording sites, blue: putative deeper recording sites). (B) Scatterplot of average delta wave trough and peak amplitudes for each recorded electrode group. **(C)** The proportion of nonsilent delta waves was not significantly different between recordings from putative deep (blue) and recordings from putative superficial (red) recording sites (dots: individual recording sessions; bars: mean \pm s.e.m, P = 0.95305, Wilcoxon rank-sum test). **(D)** Hippocampal ripple activity predicts delta spikes independently of the depth of the recording site. Correlations between hippocampal ripple activity (recorded in 200 ms windows preceding delta waves) and delta spikes were significantly greater than in time-shifted controls (grey bars) for recordings in both putative deep (blue, P = 0.028, Monte-Carlo test) and superficial (red, P = 0.028, Monte-Carlo test) and superficial and deeper recording sites (P = 0.674, Monte-Carlo test) (bars: mean \pm s.e.m).



Fig. S5. Characteristics of silent and non-silent delta waves. (A) Gamma power (z-scored; curves and shaded areas, mean \pm s.e.m.) decreased around delta waves regardless of whether spikes were detected during the delta waves (non-silent, red) or not (silent, black). (B) Ripple occurrence rate (curves and shaded areas, mean \pm s.e.m.) preceding and following delta waves was similar for silent (black) and non-silent (red) delta waves. (C) Peri-event spectrograms centered on non-silent (left) and silent (right) delta waves, normalized relative to baseline power recorded 1s before delta waves (dashed lines). Note the increase in spindle-band activity following both non-silent and silent delta waves.



Fig. S6. Delta spike detection. (A) The proportion of non-silent delta waves increases with the number of recorded neurons (Pearson's r = 0.7456, P = 0.0014; dashed line, regression line). **(B)** For each session, delta waves were repeatedly detected using decreasing random subpopulations of neurons. Color-code: total number of neurons recorded in the session (range, 4–68). Note that the proportion of delta waves with detected delta spikes is greatly underestimated in limited data sets, and this effect is even stronger in sessions with fewer recorded neurons (cold colors). **(C)** The mean number of neurons active during a given delta wave linearly increases with the number of recorded neurons (Pearson's r = 0.9041, P < 0.001; dashed line, regression line). Shaded area: 95% confidence intervals for the regression line obtained with jackknife resampling. Extrapolating to the total number of neurons in the mPFC (*34*), the number of neurons expected to fire in any given delta wave would be 3800–4600.



Fig. S7. Activity components during delta waves. (A) Examples of correlated firing during delta waves (red circles). (B) Example significant templates (putative cell assemblies) identified using independent component analysis (ICA). (C) Prediction gain (bars: mean \pm s.e.m.) of a generalized linear model (GLM) trained to predict the delta spikes of each prefrontal neuron, based on the delta spikes of all other prefrontal neurons (peer activity) for endogenous as well as induced delta waves (see Fig. 4). Prefrontal peer activity substantially improved the prediction gain compared to a control condition where delta spikes were predicted based solely on overall delta activity levels, i.e. summing spikes from all peers and ignoring cell identities, for both endogenous (P = 0.0028, Wilcoxon signed rank test) and induced delta waves (delayed stimulation: P < 0.001; coupled stimulation: P < 0.001, Wilcoxon signed rank test; endogenous vs delayed stimulation: P = 0.0221; coupled vs delayed stimulation: P < 0.001; endogenous vs delayed stimulation: P = 0.4377, Wilcoxon rank-sum tests).



Fig. S8. Hippocampal prediction of delta spikes does not critically depend on data recorded from any single rat. Cross-correlations (curves and shaded areas, mean \pm s.e.m.) between hippocampal ripple activity (sliding window) and delta spikes (fixed, 0s). Observed cross-correlations (orange) vs time-shifted control cross-correlations (grey) for hippocampal ripple activity preceding delta waves (horizontal orange line: Monte-Carlo test, P < 0.05). Each panel repeats the analysis presented in Fig. 2 leaving one rat out.



Fig. S9. Hippocampal ripple activity better predicts cortical spikes in the presence of a delta wave. (A) Cross-correlations (curves and shaded areas, mean \pm s.e.m.) between hippocampal ripple activity (fixed, 0s) and cortical spikes (sliding window), in the presence (left) or absence (right) of a delta wave following (within 200 ms) the ripple. Observed cross-correlations (orange, blue) vs time-shifted control cross-correlations (grey) (horizontal orange and blue lines: Monte-Carlo tests, P < 0.05). (B) Correlation between hippocampal ripple activity and subsequent cortical spikes ([0,200] ms time window) in the presence (orange bar) or absence (blue bar) of delta waves (grey bars, time-shifted control correlations). ***P < 0.001, Wilcoxon signed rank tests. (C) Same as (B) but using a more stringent 30 ms window centered on the delta wave following the ripple (orange) or at 130 ms post ripple (blue). ***P < 0.001, Wilcoxon signed rank tests.



Fig. S10. Hippocampal ripple activity fails to predict delta spikes in sleep preceding behavior. (A) Cross-correlations (curves and shaded areas, mean \pm s.e.m.) between hippocampal ripple activity (sliding window) and delta spikes (fixed, 0 s) in sleep before the task. Observed cross-correlations (orange) were not significantly different from time-shifted control cross-correlations (grey) (Monte-Carlo test, P > 0.05). (C) Comparative distribution between data and control in (A). There was no enrichment of positively correlated pairs of hippocampal ripple activity and subsequent delta waves in sleep before the task. (C) Performance of a GLM trained to predict prefrontal activity during delta waves in sleep before the task based on preceding hippocampal ripple activity (200 ms window), measured as percent improvement relative to a shuffled control (prediction gain). Multiple single-unit hippocampal activity did not improve performance relative to global hippocampal drive ignoring cell identity (P = 0.6251, Wilcoxon signed rank test). Neither model could significantly predict prefrontal activity during delta waves in sleep before the task (multiple single units, P = 0.3640; summed activity, P = 1; Wilcoxon rank-sum tests).



Fig. S11. Hippocampal ripple activity predicts delta spikes. (A) Cross-correlations (curves and shaded areas, mean \pm s.e.m.) between hippocampal ripple activity (sliding window) and delta spikes (fixed, 0s) separately for non-partner cells (left) vs partner cells (right). Observed cross-correlations (orange) vs time-shifted control cross-correlations (grey) for hippocampal ripple activity preceding delta waves (horizontal orange line: Monte-Carlo test, P < 0.05). Hippocampal spikes could predict delta spikes only for partner cells (right). (B) Enrichment in positive correlations (comparative distribution between data and control in (A)) when hippocampal activity was correlated to subsequent prefrontal delta spikes. (C) Performance of a GLM trained to predict prefrontal activity during delta waves based on preceding hippocampal ripple activity (200 ms window), measured as percent improvement relative to a shuffled control (prediction gain). Delta partner spikes were significantly predicted by multiple single-unit hippocampal activity (P < 0.001, Wilcoxon rank-sum tests), but not by global hippocampal drive ignoring cell identity (non-partner hippocampal units, P = 1, Wilcoxon rank-sum test). ***P < 0.001, Wilcoxon signed rank tests.



Fig. S12. Delta assemblies are uniquely expressed during task performance. Comparison of assemblies during delta waves and non-delta periods in sleep preceding behavior (pre-sleep), during behavior, and during delta waves and non-delta periods in sleep following behavior (post-sleep). (A) Comparison of post-sleep delta assemblies and all other assemblies (orange: correlation between assemblies; grey: shuffled controls). Correlations were significant only between delta assemblies and assemblies expressed during behavior. (B) Comparison of pre-sleep delta assemblies and other assemblies. No correlation was significant. (C) Comparison of non-delta assemblies. Assemblies expressed during behavior were significantly correlated with both post- and pre-sleep assemblies, consistent with similar findings about hippocampal ripple activity during sleep.



Fig. S13. Delta waves isolate cortical computations. Simultaneous recordings of prefrontal and hippocampal activity around delta waves (grey shaded rectangles). Top: proportion of prefrontal spikes predicted by the firing of hippocampal cells (partner spikes). Center: raster plot of spikes emitted by prefontal units (red ticks: partner spikes, grey ticks: other spikes). Bottom: simultaneously recorded local field potentials in the mPFC (blue: delta wave) and hippocampus (broadband and ripple-band filtered signal). Black calibration bars: 0.5 s.



Fig. S14. Selective silencing of cortical spikes during delta waves. (A) Proportion of prefrontal spikes explained by the preceding hippocampal activity (partner spikes). Observed proportions (red) were significantly different from time-shifted controls (grey), precisely during delta waves (curves and shaded areas, mean \pm s.e.m.; horizontal red line: Monte-Carlo test, P < 0.05). **(B)** Average rate (curves and shaded areas, mean \pm s.e.m.) of prefrontal partner spikes (green) and other spikes (blue) around delta waves. Note that the firing rate of partner spikes remains relatively constant, whereas the firing rate of other spikes decreases dramatically at t = 0, effectively boosting the signal-to-noise ratio during delta waves.



Fig. S15. Cortical stimulation triggers both silent and non-silent delta waves. (A) Occurrence rate (mean \pm s.e.m.) for silent (grey) and non-silent (red) delta waves following cortical stimulation. Rate is normalized by the total number of silent and non-silent delta waves, respectively (inset: non-normalized rates). (B) Proportion of delta waves following stimulation pulses within 200 ms (bars: median \pm s.e.median; open circles: individual sessions) was not different between silent (black) and non-silent (red) delta waves (P > 0.05, Wilcoxon signed rank test).

session	delta waves	mPFC units	HPC units	mPFC units with HPC partners	mPFC units with mPFC peers
1	1150	19	10	8	17
2	1665	67	10	9	67
3	794	9	10	0	0
4	482	1	9	0	0
5	381	6	13	0	5
6	1512	13	8	5	5
7	1579	8	4	4	0
8	595	5	4	0	2
9	216	1	3	0	0
10	1360	12	13	5	9
11	2046	28	6	6	27
12	358	4	14	1	0
13	1910	19	23	11	17
14	3186	10	12	1	6

Table S1. Number of units recorded from the hippocampus and prefrontal cortex during slow wave sleep. Only units recorded on channels where delta waves could be detected were included.

session	unit	delta spikes	HPC partners	mPFC peers	session	unit	delta spikes	HPC partners	mPFC peers
1	1	3	0	1					
1	2	11	0	4	:	:	:	:	:
1	3	3	0	7	2	41	51	0	16
- 1	4	25	0	6	2	42	1	0	2
1	5	20	2	6	2	43	13	0	20
1	5	12	2	4	2	44	1	0	4
1	0	15	2	4	2	15	0	0 0	0
1	1	5	0	0	2	46	74	1	17
1	8	1	0	0	2	40	74	1	
1	9	11	0	3	2	47	2	0	1
1	10	5	1	5	2	48	5	0	9
1	11	17	3	6	2	49	6	0	3
1	12	4	2	3	2	50	40	2	26
1	13	20	0	11	2	51	35	0	20
1	14	4	1	5	2	52	3	0	1
1	15	1	1	2	2	53	88	0	18
1	16	8	0	7	2	54	93	0	31
1	17	52	1	11	2	55	4	0	13
1	18	13	0	2	2	56	10	0	6
1	19	18	0	3	2	57	8	1	6
2	1	16	0	15	2	58	28	0	14
2	2	66	0	16	2	59	20	0	13
2	2	42	0	21	2	60	24	0	<u>a</u>
2	3	45	0	21	2	61	10	0	11
2	4	45	0	10	2	62	5	0	3
2	5	8	0	1	2	62	11	0	10
2	6	14	0	6	2	05	11	0	10
2	7	14	0	5	2	64	8	4	10
2	8	55	0	12	2	65	18	0	13
2	9	7	0	4	2	66	8	0	6
2	10	2	0	10	2	67	25	1	8
2	11	39	2	21	3	1	1	0	0
2	12	3	0	11	3	2	2	0	0
2	13	80	1	30	3	3	1	0	0
2	14	34	0	21	3	4	2	0	0
2	15	6	0	7	3	5	3	0	0
2	16	3	0	10	3	6	2	0	0
2	17	57	0	23	3	7	10	0	0
2	18	23	0 0	9	3	8	1	0	0
2	10	20	0	7	3	9	3	0	0
2	20	2	0	10	4	1	1	0	0
2	20	2	0	10	5	1	2	0 0	0 0
2	21	17	0	4	5	2	2	0	1
2	22	2	0	8	5	2	2	0	1
2	23	11	0	4	5	3	3	0	1
2	24	6	0	6	5	4	4	0	1
2	25	3	0	6	5	5	0	0	2
2	26	6	0	6	5	6	2	0	1
2	27	2	1	7	6	1	1	0	0
2	28	21	0	10	6	2	1	0	1
2	29	23	0	4	6	3	2	0	0
2	30	17	0	13	6	4	7	1	0
2	31	20	0	14	6	5	7	0	0
2	32	1	0	3	6	6	5	0	1
2	33	14	0	15	6	7	6	2	0
2	34	2	0	12	6	8	10	0	0
2	35	5	0	8	6	9	1	0	1
2	36	1	0	1	6	10	7	2	2
2	37	<u>-</u> 4	0	4	6	11	8	1	0
2	32	· ·	1	5	6	12	8	1	0
∠ 2	30	<u>د</u>	л Т	ן 10	6	13	4	0	1
2	29	55	0	0 10	7	1	20	ñ	0
2	40	<i>i</i>	U	2		-	20	0	0
÷	÷	÷	:	:	:	÷	:		÷

Table S2. Number of delta spikes emitted by prefrontal cortical cells during slow wave sleep.

session	unit	delta snikes	HPC	mPFC	session	unit	delta spikes	HPC partners	mPFC peers
								· .	· .
:	:	:	:	:	:	:	:	:	:
7	2	5	1	0	11	18	18	0	5
7	3	3	0	0	11	19	17	0	1
7	4	1	0	0	11	20	12	0	3
7	5	1	0	0	11	21	41	0	3
7	6	23	1	0	11	22	14	0	6
7	7	4	1	0	11	23	104	0	(
7	8	4	2	0	11	24	9	0	4
8	1	12	0	0	11	25	10	1	4
8	2	4	0	1	11	20	49	1	5
8	3	50	0	0	11	21	14	1	1
8	4	38	0	0	11	20 1	10	1	1
8	5	74	0	1	12	2	1	1	0
9	1	1	0	0	12	2	1	0	0
10	1	3	0	0	12	1	1	0	0
10	2	3	1	1	12	4	18	1	5
10	3	21	1	U F	13	2	8	0	2
10	4	4	2	C A	13	3	92	1	6
10	5	9	1	4	13	4	16	0	5
10	7	10	1	2	13	5	3	3	1
10	7 Q	1	0	1	13	6	10	0	10
10	0	3	0	1	13	7	9	2	4
10	10	15	0	1	13	8	25	0	10
10	11	15	0	4	13	9	69	1	9
10	12	4	0	2	13	10	28	0	9
11	1	27	0 0	0	13	11	27	3	7
11	2	46	0	2	13	12	6	0	0
11	3	10	0	8	13	13	7	1	3
11	4	12	0	1	13	14	21	0	3
11	5	11	0	2	13	15	14	1	4
11	6	19	0	4	13	16	7	1	3
11	7	8	0	3	13	17	9	2	6
11	8	16	0	3	13	18	45	0	7
11	9	5	1	5	13	19	30	1	0
11	10	67	0	5	14	1	157	0	2
11	11	5	0	3	14	2	83	0	2
11	12	19	0	8	14	3	93	0	2
11	13	35	1	6	14	4	84	1	3
11	14	39	0	4	14	5	39	0	0
11	15	32	1	5	14	6	65	0	1
11	16	41	1	3	14	7	24	0	0
11	17	70	0	8	14	8	48	0	0
	-		•		14	9	2	0	0
:	:	:	:	:	14	10	44	0	2

Table S2 (cont.). Number of delta spikes emitted by prefrontal cortical cells during slow wave sleep.

References and Notes

- M. Steriade, A. Nuñez, F. Amzica, A novel slow (< 1 Hz) oscillation of neocortical neurons in vivo: Depolarizing and hyperpolarizing components. *J. Neurosci.* 13, 3252–3265 (1993). doi:10.1523/JNEUROSCI.13-08-03252.1993 Medline
- H.-V. V. Ngo, T. Martinetz, J. Born, M. Mölle, Auditory closed-loop stimulation of the sleep slow oscillation enhances memory. *Neuron* 78, 545–553 (2013). doi:10.1016/j.neuron.2013.03.006 Medline
- 3. L. Marshall, H. Helgadóttir, M. Mölle, J. Born, Boosting slow oscillations during sleep potentiates memory. *Nature* 444, 610–613 (2006). doi:10.1038/nature05278 Medline
- 4. S. Chauvette, J. Seigneur, I. Timofeev, Sleep oscillations in the thalamocortical system induce long-term neuronal plasticity. *Neuron* 75, 1105–1113 (2012). <u>doi:10.1016/j.neuron.2012.08.034</u> Medline
- N. Maingret, G. Girardeau, R. Todorova, M. Goutierre, M. Zugaro, Hippocampo-cortical coupling mediates memory consolidation during sleep. *Nat. Neurosci.* 19, 959–964 (2016). doi:10.1038/nn.4304 Medline
- 6. A. Sirota, G. Buzsáki, Interaction between neocortical and hippocampal networks via slow oscillations. *Thalamus Relat. Syst.* **3**, 245–259 (2005). <u>doi:10.1017/S1472928807000258</u> <u>Medline</u>
- A. Sirota, J. Csicsvari, D. Buhl, G. Buzsáki, Communication between neocortex and hippocampus during sleep in rodents. *Proc. Natl. Acad. Sci. U.S.A.* 100, 2065–2069 (2003). doi:10.1073/pnas.0437938100 Medline
- G. Girardeau, K. Benchenane, S. I. Wiener, G. Buzsáki, M. B. Zugaro, Selective suppression of hippocampal ripples impairs spatial memory. *Nat. Neurosci.* 12, 1222–1223 (2009). doi:10.1038/nn.2384 Medline
- 9. V. Ego-Stengel, M. A. Wilson, Disruption of ripple-associated hippocampal activity during rest impairs spatial learning in the rat. *Hippocampus* **20**, 1–10 (2010). <u>Medline</u>
- 10. A. K. Lee, M. A. Wilson, Memory of sequential experience in the hippocampus during slow wave sleep. *Neuron* 36, 1183–1194 (2002). <u>doi:10.1016/S0896-6273(02)01096-6</u> <u>Medline</u>
- 11. D. Ji, M. A. Wilson, Coordinated memory replay in the visual cortex and hippocampus during sleep. *Nat. Neurosci.* **10**, 100–107 (2007). <u>doi:10.1038/nn1825</u> <u>Medline</u>
- G. Rothschild, E. Eban, L. M. Frank, A cortical-hippocampal-cortical loop of information processing during memory consolidation. *Nat. Neurosci.* 20, 251–259 (2017). doi:10.1038/nn.4457 Medline
- C. M. Wierzynski, E. V. Lubenov, M. Gu, A. G. Siapas, State-dependent spike-timing relationships between hippocampal and prefrontal circuits during sleep. *Neuron* 61, 587– 596 (2009). doi:10.1016/j.neuron.2009.01.011 Medline
- A. Peyrache, M. Khamassi, K. Benchenane, S. I. Wiener, F. P. Battaglia, Replay of rulelearning related neural patterns in the prefrontal cortex during sleep. *Nat. Neurosci.* 12, 919–926 (2009). doi:10.1038/nn.2337 Medline

- P. B. Kruskal, L. Li, J. N. MacLean, Circuit reactivation dynamically regulates synaptic plasticity in neocortex. *Nat. Commun.* 4, 2574 (2013). doi:10.1038/ncomms3574 Medline
- 16. T. Gulati, L. Guo, D. S. Ramanathan, A. Bodepudi, K. Ganguly, Neural reactivations during sleep determine network credit assignment. *Nat. Neurosci.* 20, 1277–1284 (2017). doi:10.1038/nn.4601 Medline
- 17. A. G. Siapas, M. A. Wilson, Coordinated interactions between hippocampal ripples and cortical spindles during slow-wave sleep. *Neuron* 21, 1123–1128 (1998). doi:10.1016/S0896-6273(00)80629-7 Medline
- M. Rosanova, D. Ulrich, Pattern-specific associative long-term potentiation induced by a sleep spindle-related spike train. J. Neurosci. 25, 9398–9405 (2005). doi:10.1523/JNEUROSCI.2149-05.2005 Medline
- J. Seibt, C. J. Richard, J. Sigl-Glöckner, N. Takahashi, D. I. Kaplan, G. Doron, D. de Limoges, C. Bocklisch, M. E. Larkum, Cortical dendritic activity correlates with spindlerich oscillations during sleep in rodents. *Nat. Commun.* 8, 684 (2017). <u>doi:10.1038/s41467-017-00735-w Medline</u>
- 20. D. Marr, Simple memory: A theory for archicortex. *Philos. Trans. R. Soc. London Ser. B* **262**, 23–81 (1971). <u>doi:10.1098/rstb.1971.0078</u> <u>Medline</u>
- 21. G. Buzsáki, Two-stage model of memory trace formation: A role for "noisy" brain states. *Neuroscience* **31**, 551–570 (1989). doi:10.1016/0306-4522(89)90423-5 Medline
- 22. P. W. Frankland, B. Bontempi, The organization of recent and remote memories. *Nat. Rev. Neurosci.* **6**, 119–130 (2005). <u>doi:10.1038/nrn1607</u> <u>Medline</u>
- V. Lopes-dos Santos, S. Ribeiro, A. B. L. Tort, Detecting cell assemblies in large neuronal populations. *J. Neurosci. Methods* 220, 149–166 (2013). doi:10.1016/j.jneumeth.2013.04.010 Medline
- 24. K. D. Harris, J. Csicsvari, H. Hirase, G. Dragoi, G. Buzsáki, Organization of cell assemblies in the hippocampus. *Nature* **424**, 552–556 (2003). <u>doi:10.1038/nature01834</u> <u>Medline</u>
- 25. M. Steriade, I. Timofeev, Neuronal plasticity in thalamocortical networks during sleep and waking oscillations. *Neuron* **37**, 563–576 (2003). <u>doi:10.1016/S0896-6273(03)00065-5</u> <u>Medline</u>
- 26. A. Luczak, P. Barthó, S. L. Marguet, G. Buzsáki, K. D. Harris, Sequential structure of neocortical spontaneous activity in vivo. *Proc. Natl. Acad. Sci. U.S.A.* **104**, 347–352 (2007). doi:10.1073/pnas.0605643104 Medline
- L. A. Johnson, D. R. Euston, M. Tatsuno, B. L. McNaughton, Stored-trace reactivation in rat prefrontal cortex is correlated with down-to-up state fluctuation density. *J. Neurosci.* 30, 2650–2661 (2010). doi:10.1523/JNEUROSCI.1617-09.2010 Medline
- 28. J. Csicsvari, D. A. Henze, B. Jamieson, K. D. Harris, A. Sirota, P. Barthó, K. D. Wise, G. Buzsáki, Massively parallel recording of unit and local field potentials with silicon-based electrodes. J. Neurophysiol. 90, 1314–1323 (2003). doi:10.1152/jn.00116.2003 Medline

- 29. D. Jercog, A. Roxin, P. Barthó, A. Luczak, A. Compte, J. de la Rocha, UP-DOWN cortical dynamics reflect state transitions in a bistable network. *eLife* 6, e22425 (2017). doi:10.7554/eLife.22425 Medline
- N. Maingret, R. Todorova, M. Zugaro, Recordings and timed stimulation of rat dorsal hippocampal area CA1 and medial prefrontal cortex during behavior and sleep. CRCNS (2019); <u>https://doi.org/10.6080/K0TD9VJG.</u>
- L. Hazan, M. Zugaro, G. Buzsáki, Klusters, NeuroScope, NDManager: A free software suite for neurophysiological data processing and visualization. J. Neurosci. Methods 155, 207– 216 (2006). doi:10.1016/j.jneumeth.2006.01.017 Medline
- P. Barthó, H. Hirase, L. Monconduit, M. Zugaro, K. D. Harris, G. Buzsáki, Characterization of neocortical principal cells and interneurons by network interactions and extracellular features. J. Neurophysiol. 92, 600–608 (2004). doi:10.1152/jn.01170.2003 Medline
- 33. G. M. van de Ven, S. Trouche, C. G. McNamara, K. Allen, D. Dupret, Hippocampal Offline Reactivation Consolidates Recently Formed Cell Assembly Patterns during Sharp Wave-Ripples. *Neuron* 92, 968–974 (2016). doi:10.1016/j.neuron.2016.10.020 Medline
- 34. J. Willing, J. M. Juraska, The timing of neuronal loss across adolescence in the medial prefrontal cortex of male and female rats. *Neuroscience* **301**, 268–275 (2015). doi:10.1016/j.neuroscience.2015.05.073 Medline